

Allelopathic Control Effect of Complex Formulation of Compound Microbial Community and Bio-Organic Fertilizer on *Cucumber fusarium* Wilt

LIJIN QIN¹*, ZENGMING ZHONG², DANDAN WANG¹, HAIQI HU¹, DUO LI¹, YINBIAO GAO¹, SHOUZHENG WANG¹

¹Chifeng University, Chifeng 024000, China

² Beijing Qigao Biologics Co., Ltd., Beijing 100193, China

Abstract: To study the influence of complex formulation of "WoFengKang" compound microbial community and bio-organic fertilizer on allelopathic effect of cucumber fusarium wilt and field growth traits, in this experiment, different indoor treatments were taken to cultivate cucumber fusarium wilt using soil leaching liquor and Cucumis sativus growth index, fusarium wilt disease index were determined, so that allelopathic control effect of different dosages of compound microbial community on cucumber fusarium wilt can be investigated. The results showed that: compared with conventional CK, cucumber fusarium wilt cultured in different soil leaching liquor had smaller diameter. Where, colony diameter in treatment 3 was the smallest, which was 36.72 mm. Compound microbial community had certain allelopathic effect on cucumber fusarium wilt, and inhibition effect was shown with allelopathic effect reaching 27.13~43.91%. The above differently-treated soil was loaded to seedling-raising pot for Cucumis sativus planting. Cucumber fusarium wilt (FOC) was inoculated in the rough leaf stage of Cucumis sativus, thus reducing the disease index of cucumber fusarium wilt with obvious control effect reaching 55.94~72.63%. Where, treatment 3 demonstrated the best effect and allelopathic control effect reached 72.63% after 15d inoculation. The complex formulation of the two promoted vegetative growth of Cucumis sativus, lowered node of the first female flower bud, increased the number of female flowers within 30 nodes, so that 3.25d early flowering and 4.75d delayed seedling raising were achieved. Therefore, application of microbial community amid and after Cucumis sativus field planting has certain allelopathic control effect on cucumber fusarium wilt, which can effectively reduce the incidence of cucumber fusarium wilt and provide a scientific basis for the allelopathic prevention and control of soil-borne diseases in Cucumis sativus.

Keywords: compound microbial community; cucumber fusarium wilt; soil leaching liquor; allelopathic effect; prevention and control effect

1.Introduction

Cucumber fusarium wilt, a soil-borne disease with serious hazard in *Cucumber* production and cultivation, is difficult to control. So far, there has been no effective control method yet. At present, chemical pesticides are mainly used for its prevention and control in production of *Cucumber*. However, the control effect is poor and environmental pollution and pesticide residues are easy occurrences [1, 2]. Compound microbial community is one or several kinds of beneficial fungus that can move freely in the soil or serve as epiphyte in the plant rhizosphere, root surface, stems and leaves, etc., which can promote plant growth. Allelopathic effect means favorable or unfavorable effect of metabolic secretions of plant (or including microorganisms) on other plants (including microorganisms) in the environment [1-15].

Understanding allelopathic effects of different plants and using this allelopathic promotion or inhibition between plants (including microorganisms) to control soil-borne diseases of plant is a prominent problem waiting urgent solution in today's "scale" and "simplified" planting production.

^{*}email: lijin.q612@gmail.com



Studies have confirmed that compound microbial community can inhibit the reproduction of pathogenic microorganisms and effectively prevent and control the occurrence of plant diseases. Yang et al. [16], measured the spores and hyphae of *Botrytis cinerea* using flat plate confrontation method, finding that 86.25% control rate of cucumber gray mold was achieved with strong disease prevention effect shown when microbial fermentation broth had a matching of 5:4:1 by volume and the number of living spores was 1.5×10^7 cfu/mL. Ji et al. [17] confirmed that the application of compound microbial community reduced the soil nematode propagation index, with control rate of root-knot nematodes reaching 56.5 and 42.5%, respectively.

In recent years, scholars have studied the allelopathic effect of plants and its rhizosphere biological mechanisms, confirming that both positive and negative effects of interactions between plants of the same or different species are a result of the interaction between root exudates-mediated plants and specific microorganisms. Root exudates play an important role in regulating the structure and function of rhizosphere micro-ecological systems, and their interaction with specific microorganisms in the rhizosphere constitutes an important process in allelopathic effect of crops [18-28]. At present, existing researches on compound microbial community in China are concentrated on pot experiments. Whether there is allelopathic effect on cucumber fusarium wilt when compound microbial community is used in *Cucumis sativus* production, how about the allelopathic effect, how it affect *Cucumber* growth and biomass production, further research is needed in these regards [17, 39-33].

In this experiment, bio-organic fertilizer was used as *Cucumber* base fertilizer, compound microbial community was applied to *Cucumis sativus* in different growth stages at the later stage, and the two were compounded. Through culture of colony diameter of cucumber fusarium wilt, investigation of incidence of field fusarium wilt and recording and determination of vegetative growth traits of *Cucumber* by laboratory, influence of complex formulation of "WoFengKang" bio-organic fertilizer and compound microbial community on allelopathic effect of cucumber fusarium wilt, field allelopathic control and field growth traits of *Cucumis sativus* was investigated, which will provide a scientific basis for allelopathic prevention and control of soil-borne diseases and high-efficiency and high-quality cultivation of greenhouse vegetables in the future.

2. Material and methods

2.1. Overview of the test site

The test site is located in Hongshan District, Chifeng City, Inner Mongolia (118°53′E, 42°13′N). The soil in this area is clay loam. The annual average temperature is 7, the annual illumination hour is 3000 h, the frost-free period is 135~145 d, and the annual average precipitation is 370 mm, and the site is in temperate semi-arid continental monsoon climate zone. The basic chemical properties of the tested soil at 0~20 cm were: pH 8.39, conductivity 93.65 μ S/cm, organic matter 1.34%, nitrate nitrogen 7.28 mg/kg, ammonium nitrogen 0.11 mg/kg, available phosphorus 134.5 mg/kg, and available potassium 106.2 mg/kg.

2.2. Test materials

'Jinchun No. 4' *Cucumis sativus* variety: provided by Tianjin Kerun Cucumber Research Institute; Cucumber fusarium wilt: provided by the Vegetable Cultivation Laboratory of the Agricultural College of Inner Mongolia Agricultural University; "WoFengKang" bio-organic fertilizer: provided by Beijing Qigao Biologics Co., Ltd., total nutrient 5%, organic matter 45%, effective viable count 50 million/ gram, containing trace elements like calcium, magnesium, sulfur, boron, zinc , molybdenum, copper, iron, net content 40kg/bag; "WoFengKang" compound microbial community: provided by Beijing Qigao Biologics Co., Ltd., effective viable count 5 billion/ml, water agent, 1000 ml/bottle, effective strains are *Bacillus subtilis* and *Clonostachys rosea*;

Conventional control organic fertilizer: provided by Inner Mongolia Liaozhongjing Agricultural Science and Technology Co., Ltd., total nutrient 5%, organic matter 45%, humic acid 15%, amino acid 1.5%, effective viable count 200 million / gram, CaO 26%, SiO 26%, net content 40kg/bag;



Mengduoli Sanan compound fertilizer: provided by Neimenggu Zhongjing Huier Biological Technology Co., Ltd., total nutrient≥54%, N:P₂O₅:K₂O =18:18:18, net content 40kg/bag;

Rare earth superphosphate fertilizer: provided by Qinhuangdao Tianhao Chemical Co., Ltd., Ca \geq 17%, S \geq 11%, effective phosphorus pentoxide \geq 12%, rare earth content 0.2%-0.3%, net content 40kg/bag.

2.3. Experimental design

This experiment took the conventional fertilization by farmers in Chifeng area as the control, and used "WoFengKang" bio-organic fertilizer as the base fertilizer which was compounded with "WoFengKang" compound microbial community in later stage. There were 5 treatments, each treatment had 3 repetitions with regular field management. The experiment was completed in the cold shed of the research base of Chifeng University from May to September 2019. "WoFengKang" bio-organic fertilizer was applied in the amount of 3, 6, 9, and 12 bags/mu. The conventional CK was applied according to the local farmers' habit of 9 bags/mu. *Cucumis sativus* was sowed, followed by seedling raising on May 29, 2019 and planted on July 15. The plant spacing was 35 cm×55 cm, and the plot area was 5.5 m². The specific treatment combination and fertilization scheme are shown in Tables 1 and 2.

 Table 1. Complex formulation combinations of "WoFengKang" bio-organic fertilizer

 and compound microbial community

Different treatments	"WoFengKang" bio- organic fertilizer (kg/mu)	"WoFengKang" compound microbial community (bottle/mu)	Organic fertilizer (kg/bag)	Mengduoli Sanan compound fertilizer (kg/mu)	Rare earth superphosphate fertilizer (kg/mu)	
Treatment 1	360	2	0	40	40	
Treatment 2	360	4	0	40	40	
Treatment 3	360	6	0	40	40	
Treatment 4	360	8	0	40	40	
Conventional CK	0	0	360	40	40	

Table 2. Period of use and dosage of "WoFengKang" compound microbial community

Different treatments	<i>Cucumis</i> <i>sativus</i> planting period (times)	Rejuvenation period (times)	Blossom and fruit period(times)	First fruiting period(times)	Full bearing period(times)	7d before seedling raising (times)
Treatment 1	1	1	0	0	0	0
Treatment 2	1	1	1	0	1	0
Treatment 3	1	1	1	1	1	1
Treatment 4	1	1	1	2	2	1
Conventional	0	0	0	0	0	0

Note: "WoFengKang" compound microbial community was used from the time of *Cucumber* planting, with a dosage of 1 bottle/mu/time. The dosages are the same for the following. It was used once in the planting period, once after rejuvenation, once in the blossom and fruit period, 1-2 times in the initial fruiting period, 1-2 times in the full bearing period, and once at 7d before seedling raising.

2.4 Measurement indicators and methods

2.4.1 Method to determine colony diameter and allelopathic effect

Amid seedling raising of the *Cucumis sativus* (October 20, 2019), the above differently- treated soil samples were taken and the soil at 0~20 cm layer of rhizosphere of the *Cucumis sativus* plants was collected using a soil drill. By "W" type 5-point sampling method, the soil was thoroughly mixed and loaded in plastic bag, sent to the indoor laboratory. Fresh soil samples of different treatments were extracted in the laboratory using ethanol extractant to obtain differently- treated soil leaching liquor.



Under aseptic conditions, PDA medium was added to the above soil leaching liquor to culture cucumber fusarium wilt in a constant temperature drying oven at 28°C and measure the colony diameter and allelopathic effect.

Preparation of soil leaching liquor: Using ethanol as a soil extractant, the extraction was performed according to soil sample mass: extractant volume = 1:2. After oscillation extraction for 24 h on a shaker (25° C, 110 r/min), the soil was filtered through 6 layers of gauze and then filtered through a layer of qualitative filter paper to obtain the soil leaching liquor.

Plate preparation: 2 mL of the above differently-treated soil leaching liquor was transferred to a 9 cm diameter Petri dish, then 18 mL molten PDA medium was added and mixed to prepare a plate. 2 mL soil leaching liquor was added as treatment, while 2 mL ethanol extractant plate and 20 mL PDA medium (blank) plate were added as ethanol CK and total CK, respectively.

Inoculation culture: Under aseptic conditions, a fungus cake was punched with a 0.6 cm-diameter puncher, and transferred to the above different mediums using inoculating needle, with the hyphae downward and one cake per dish, which was placed in the center of the dish. It was then cultured in a 25°C constant temperature incubator, and each treatment was repeated 5 times. The colony diameter was determined by crossing method.

Colony diameter (cm) = average value of the measured colony diameter -0.6; Allelopathic effect (%) = (control colony diameter -treated colony diameter) / control colony diameter $\times 100$.

2.4.2. Field Inoculation and determination of *Cucumber fusarium* wilt

During *Cucumis sativus* seedling raising (October 20, 2019), the above differently-treated soils were loaded to seedling-raising pot for *Cucumis sativus* planting. Cucumber fusarium wilt (FOC) was inoculated when the rough leaf of *Cucumis sativus* fully unfolded and incidence of cucumber fusarium wilt was recorded after inoculation for 7d. Then, measure the disease index and prevention and treatment effects. There were 25 seedling raising pots in each treatment, which were 8 cm in diameter and filled with soil. Each treatment was repeated 4 times. *Cucumis sativus* seeds were germinated at a constant temperature (28°C) illumination incubator. After germination, the seeds were sown in a seedling-raising pot loaded with soil, one seed per hole, which were then routinely managed. When the rough leaf of *Cucumis sativus*, and cucumber fusarium wilt was inoculated by root pouring (FOC, 1.0×10^6 in concentration, 5mL was poured to the square per pot). The incidence of the disease was investigated after 7d of inoculation. Investigation was made once every other day for a total of 6 times. The disease index and control effect were measured. The reference criteria for different grades of cucumber fusarium wilt are as follows [34]:

Grade 0, no symptom;

Grade 1, the hypocotyls and cotyledons are slightly wilted, and the cotyledons lose luster;

Grade 2, plant has mild wilting or there is cotyledon wilting;

Grade 3, the plant is obviously wilted or dwarfed;

Grade 4, the plants are severely wilting or withering.

Disease index = Σ (condition level × number of diseased plants at this level) / (the highest level of disease × total number of plants) × 100%;

Control effect (%) = (control disease index – treated-disease index) / control disease index \times 100%.

2.4.3 Recording and determination of *Cucumis sativus* field growth traits

After the first female flower of *Cucumis sativus* showed initial fruit expansion, the initial flowering stage and the fruiting stage, the first female flower node order, the number of female flowers in $1\sim30$ nodes, the leaf color and the fruit color were recorded. Field vegetative growth index of *Cucumis sativus* was measured in the early and middle stages of fruiting, including *Cucumis sativus* stem length, stem diameter, the maximum leaf area and the number of leaves. 15 plants were labeled at the three positions (front, middle and back) in each zone under each treatment, and the average of all the



data was calculated [35-40].

Initial flowering period: also known as budding period or vine growth stage, which is from planting to fruit expansion of first female flower;

Fruiting period: from fruit expansion of the first female flower to seedling raising;

Leaf color: observed in the initial flowering stage and the fruiting stage, generally divided into light green, green and dark green;

Fruit color: observed during maturity, divided into three color uniformity levels of deep, medium and light according to the characteristics of the variety;

Stem length: the highest plant height in whorl stage is measured by a tape measure;

Stem diameter: 1 cm from the earth surface as measured by a vernier caliper;

Maximum leaf area: select the largest leaf of the plant, measure the length and width with a tape measure, and calculate the product of the two;

Number of leaves: a leaf with a diameter of ≥ 5 cm;

The first female flower node and the number of female flowers within 1~30 nodes of the above labeled plants were recorded.

2.5. Data processing

Analysis of variance and data processing were performed using Microsoft Excel 2007 software, SPSS 17.0 software and SAS software [40-42].

3. Results and discussions

3.1 Effects of complex formulation on colony diameter and allelopathic effect of *Cucumber* fusarium wilt

The colony diameter of cucumber fusarium wilt cultured by ethanol extract with different complex formulations of compound microbial community and bio-organic fertilizer (Figure 1). Compared with conventional CK, ethanol CK and total CK, complex formulation of the two results in smaller colony diameter, and as more compound microbial community is used, colony diameter gradually decreases until it no longer changes when the dosage increases to a certain extent. This indicates that the use of compound microbial community has a certain allelopathic effect on cucumber fusarium wilt, and this effect is inhibitory (P<0.05). Where, treatment 3 had the smallest colony diameter of 36.72 mm, which was 43.91%, 40.56% and 53.48% lower than that of conventional CK, ethanol CK and total CK, respectively.

Compared with conventional CK, the 4 treatments with compound microbial community show allelopathic effects (Figure 2). Different compound microbial community treatments have obvious allelopathic effects compared with conventional CK (P < 0.05), and conventional CK has allelopathic effect of 27.13%~43.91%, indicating that compound microbial community has obvious allelopathic inhibition on cucumber fusarium wilt, and the allelopathic inhibition effect is gradually strengthened with the increase in concentration of compound microbial community, but after a certain concentration is reached, the effect is gradually weakened.



Treatment 1 Treatment 2 Treatment 3 Treatment 4





Conventional CK Ethanol CK Total CK **Figure 1.** Colony diameter of cucumber fusarium wilt cultured in PDA medium treated with different ethanol extracts at 25°C for 144 h



Figure 2. Allelopathic effects of conventional control on *Cucumber fusarium* wilt in different treatments

3.2. Control effect of complex formulation on *Cucumber fusarium* wilt in field and pot experiments

The soils in different complex formulations were loaded to nutrition pot for *Cucumis sativus* planting. Cucumber fusarium wilt (FOC) was inoculated using root pouring when the rough leaf of *Cucumis sativus* fully unfolded and incidence of cucumber fusarium wilt was recorded after inoculation for 7 d. Then, measure the disease index and prevention and treatment effects, with results shown in Table 3. Cucumber fusarium wilt under treatments with compound microbial community has significantly lower disease index than conventional CK (P< 0.05). With the delay of the investigation period, the disease index of each treatment shows an increasing trend, which stabilizes after 13d inoculation. The control rate of different treatments reaches 55.94~72.63% compared to conventional CK.

	7d after inoculation		9d after inoculation		11d after inoculation		13d after inoculation		15d after inoculation	
Different treatments	Disease index	Control effect (%)	Disease index	Control effect (%)	Disease index	Control effect (%)	Disease index	Control effect (%)	Disease index	Control effect (%)
Treatment1	17.86b	55.94	20.26b	57.12	21.27b	59.48	27.17b	60.78	28.71b	61.70
Treatment2	16.69b	58.83	19.75b	63.59	22.79b	63.46	26.82b	61.29	27.29b	65.17
Treatment3	13.57c	66.53	16.67c	69.27	17.59c	71.80	19.38c	72.03	19.69c	72.63
Treatment4	15.41b	61.99	18.78b	65.38	19.56c	68.64	21.49c	68.98	24.98b	69.77
Conventional CK	40.54a		54.25a		62.37a		69.28a		69.74a	

Table 3. The incidence and control of cucumber fusarium wilt under different treatments

Note: Different lowercase letters indicate significant differences between the same index at the 0.05 level. The same below.



3.3. Effect of complex formulation on field growth and development of *Cucumis sativus* **3.3.1. Effect on the field growth traits of** *Cucumis Sativus*

Field growth traits of *Cucumis sativus* under different treatments are shown in Table 4. This table indicates that the initial flowering period of *Cucumis sativus* treated with compound microbial community was 3.25d earlier than that of conventional CK, while the fruiting period is delayed by 4.75d. *Cucumis sativus* plants have deeper leaf color than conventional CK, the color under compound microbial community is green and dark green, while that of conventional CK is light green. *Cucumis sativus* fruit has uniform color that is deeper than that of conventional CK. Significant difference (P < 0.05) is achieved between compound microbial community treatment and conventional CK in terms of stem length, stem diameter, maximum leaf area and leaf number, indicating increase by 7.43%~20.14%, 10.77%~52.67%, 3.61%~42.38% and 15.83%~32.35%, respectively compared with conventional CK. Complex formation of the two can promote early flowering and prolong the fruiting period of *Cucumis sativus*, which lays a solid foundation for high and stable yield of *Cucumis sativus*.

Table 4. Survey of field grow	th traits of Cucumis sativus.
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			2	0				
Different treatment	Initial flowering period(DD/MM) p	Fruiting period(DD/MM)	Leaf color	Fruit color	Stem length	Stem diameter (mm)	Maximum leaf area(cm ²)	Number of leaves
Treatment 1	17/8-20/8	14/10-16/10	Green	Medium, uniform	144.25±9.62c	10.39±0.64b	559.78±41.17b	16.83±1.30a
Treatment 2	16/8-19/8	15/10-16/10	Green	Medium, uniform	153.12±11.15b	11.31±0.71b	564.79±39.25b	17.18±1.31a
Treatment 3	15/8-18/8	18/10-20/10	Dark green	Dark, uniform	187.21±13.45a	14.32±0.99a	769.23±53.53a	19.23±1.66a
Treatment 4	15/8-18/8	16/10-18/10	Dark green	Dark, uniform	177.87±12.60a	13.34±0.47a	$676.58{\pm}47.37a$	18.80±1.48a
Conventional control	19/8-22/8	11/10-13/10	Light green	Light, uneven	155.83±12.97b	9.38±0.36b	540.28±37.79b	14.53±1.07b

3.3.2. Effect on *Cucumis sativus* flowering

Different treatments lower the first female flower node order of *Cucumis sativus* (Figure 3). This is indicates that complex formation of the two treatments lowers the first female flower node order of *Cucumis sativus*, and the average node order is 4.19, which is significantly lower than conventional CK by $11.20\% \sim 24.56\%$ (*P*< 0.05). It suggests that the use of compound microbial community and bio-organic fertilizer can lower the first female flower node order of *Cucumis sativus* and promote early flowering and fruiting of *Cucumis sativus*.

The effect of different treatments on the number of female flowers per 5 nodes of 1~30 *Cucumis sativus* nodes is shown in Figure 4, that shows the different treatments increase the number of female flowers per 5 nodes of 1~30 *Cucumis sativus* nodes. Compared with conventional CK, insignificant difference exists in the number of female flowers per 5 nodes of the first 15 nodes, while there are significant differences in female flower number per 5 nodes after the 15th node (P< 0.05). The number of female flowers in 1~30 nodes of *Cucumis sativus* treated with different complex formulations increases by 1.73~3.73 nodes compared to conventional CK, respectively, with an increase of 14.98%~32.29%. The increase in female flower number in 30 *Cucumis sativus* nodes is favorable to early yield formation of *Cucumis sativus*, which lays a foundation for achieving higher economic benefits of *Cucumis sativus*.



Figure 3. Effect of complex formulation of compound microbial community and bio-organic fertilizer on the first female flower node order of *Cucumis sativus*



node order

Figure 4. Effect of formulation of compound microbial community and bio-organic fertilizer on the number of female flowers per 5 nodes of 1~30 *Cucumis sativus* nodes

Colony diameter is often used as an indicator for measuring colony growth of pathogenic colonies. Studies on the allelopathic effects of plants and its soil leaching liquor on pathogenic bacteria have been reported. Liu et al. [43] proved that potassium chloride-treated soil leaching liquor had a significant inhibitory effect on the growth of *Fusarium graminearum*; Huang et al. [44] found that leaching liquor of *Aubergine* root has significant promoting effect on the growth of three pathogens of *Verticillium dahlia*, *Sclerotinia sclerotiorum* and *Botrytis cinerea*. The members of Ph.D. thesis group have reported the allelopathic effects of leaching liquor of Rhizosphere zone fresh and rot roots of parsley on cucumber fusarium wilt [45-47]. Qin et al. [34] found that the soil leaching liquor of intercropped parsley and *Cucumis sativus* had an allelopathic effect on cucumber fusarium wilt, and as the intercrop released more allelopathic matters, the colony reproduction and growth of pathogen in cucumber fusarium wilt would be significantly inhibited. Among the three types of soil leaching liquors, ethanol colony had the smallest diameter. At present, it has been confirmed that allelochemicals isolated from crop root exudates mainly include organic acids and its derivatives [48], esters [49], phenols [50] and sulfides [51]. Moreover, studies have shown that allelopathic inhibition of *Brassicaceae* and *Bassica* family plants on bacteria and *Rhizoctonia solani* in soil mainly concerns the



three nitrogen-containing compounds secreted by the roots [52]. Gao et al. [53] confirmed that ethanol extract of fresh parsley root can secrete four nitrogen-containing compounds. After complex formulation of the two, what substances are produced in *Cucumis sativus* rhizosphere? Do these substances have allelopathic effect? What is the role and function of allelochemicals? These remains to be further studied.

Yang et al. [54] found that aqueous leaching liquor of parsley root with a higher concentration has an inhibition rate of $11.05 \sim 29.31\%$ against *Fusarium oxysporum*. Han [45] confirmed that the highest inhibition rate of parsley against cucumber fusarium wilt was 94.37%. Qin et al. [34] confirmed that the root exudates of parsley have certain inhibitory effect on cucumber fusarium wilt, and the inhibitory effect was obvious. The maximum inhibition rate against cucumber fusarium wilt was up to 75.90% with the delay of crop growth period.

At present, there have been researches on controlling plant diseases using microbial community. Zhang et al. [55] irrigated root using microbial community (1kg/mu) + amino acid (1kg/mu) for 5 consecutive times during the whole growth period of bananas, finding that damage of banana wilt was lowered, disease index was lower, which favored normal growth of banana plants. Deng et al. [56] confirmed that the application of functional compound microbial communities and bio-organic fertilizers had a control rate of 68.23% against root-knot nematode in Radish rhizosphere. Lu et al. [57] found that different ratios of soil conditioner and compound microbial communities reduced the incidence of tobacco bacterial wilt, with the highest disease control rate up to 89.99%. There has been no report on the inhibition of cucumber fusarium wilt by topdressing and drip irrigation of compound microbial community. In this study, the differently-treated soils were loaded to seedling-raising pot for Cucumis sativus planting and cucumber fusarium wilt (FOC) was inoculated by root pouring at the seedling stage. The study found that differently- treated soils showed certain allelopathic inhibition on cucumber fusarium wilt, significantly reducing the field incidence of cucumber fusarium wilt. Chen et al. [58] confirmed that ethanol extract of parsley has a strong inhibitory effect on Foc growth. Gao et al. [59] confirmed that five organic acids can be separated from the ethanol extract of fresh parsley. Where, 4-methoxysalicylic acid belongs to salicylic acid, and the salicylic acid signaling pathway in plants is closely related to disease resistance of plants [60]. Therefore, GC-MS identification of allelochemicals in the soil with complex formulation of the two is the next work to be done.

In order to deal with the current soil acidification, hardening, secondary salinization and frequent soil-borne diseases in China, many biological companies and research institutes have developed and prepared different functional antibiological inoculants which regulate the rhizosphere microbial flora to achieve dominance of beneficial bacteria, so that the purposes of disease control, production and quality increase, soil improvement can be reached. The "WoFengKang" compound microbial community used in this experiment was jointly developed by the Chinese Academy of Agricultural Sciences and Beijing Qigao Biologics Co., Ltd. The effective strains were Bacillus subtilis and Clonostachys rosea. For its mechanism, on the one hand, diseases are suppressed through competition with harmful bacteria for nutrition and living space. Secondly, growth of pathogenic bacteria is inhibited by producing substances such as antibacterial peptides, chitinase and antibiotics, and also, the produced auxins, cytokinins and indole acetic acid can hasten rooting and improve plant disease resistance. Studies have shown that compound microbial community can promote the growth of tomato [61], pepper [62], strawberry [63], amaranth [64], potato [65]. The effect of compound microbial community on *Cucumis sativus* growth has not been reported. Fu Ruimin [77] reported the application of compound microbial community on melons, finding that microbial agents-treated melons have thicker, larger and greener leaves and more vigorous growth than conventional controls. It is also more evident that compound can affect on some cellular changes [67-77].

4. Conclusions

This study shows that: complex formulation of compound microbial communities and bio-organic fertilizers at different concentrations can promote vegetative growth of *Cucumis sativus*, advance



flowering and fruiting, lower the first female flower node order, increase the number of female flowers within 30 nodes, thus laying a good foundation for premature and high yield of *Cucumis sativus*, so that economic benefit is higher. This may be because beneficial bacteria of the compound microbial community produces some plant growth hormone through its physiological metabolic activity, which promotes plant rooting and hastens the growth of plant shoots. However, can these plant growth hormones be produced? What substances are these hormones? Further testing is needed. Studies have confirmed that allelochemicals secreted by plant roots can induce population effects of specific microorganisms in the rhizosphere, causing explosive growth of these microbes and ultimately affecting the growth of target plants [18], which are consistent with the results of this study.

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